INCORPORATION OF DOUBLY LABELLED SODIUM ACETATE- 13C2 INTO PHYTUBERIN AND OTHER SESQUITERPENES
IN POTATOES: EXPERIMENTAL CONFIRMATION OF POSTULATED C-C CLEAVAGES

Albert Stoessl and Edmund W.B. Ward

Agriculture Canada Research Institute, University Sub Post Office London N6A 5B7, Canada

J.B. Stothers

Department of Chemistry, University of Western Ontario, London, Canada (Received in USA 7 June 1976; received in UK for publication 26 July 1976)

In a continuing study of the sesquiterpenoidal stress metabolites of the <u>Solanaceae</u>, we fed sodium  $(1,2^{-13}\text{C})$ -acetate  $(90\%^{-13}\text{C},1\text{ g})$  to potatoes (120 lb) which had been inoculated 18 h previously with a spore suspension of <u>Monilinia fructicola</u>. After a further 30 h incubation, chromatography of the ether-soluble material from the diffusates afforded lubimin (1,16 mg) and hydroxylubimin (2,7.5 mg); both 1 and 2 showed the same labelling patterns in their  $^{13}\text{C}$  n.m.r. spectra as those found for samples previously isolated from the interaction of the fungus with <u>Datura stramonium</u>. 1 Rishitin  $^{2}$  (3) was obtained as the major metabolite (<u>ca</u>. 70 mg) and its  $^{13}\text{C}$ 

n.m.r. spectrum (Fig. 1) revealed a labelling pattern consistent with the postulated loss of C-15. $^{3,4}$  Disappointingly, the biosynthetically most interesting compound, phytuberin $^{5,6}$  (4), was isolated in very small amount (< 1 mg) but some information was obtained from its  $^{13}$ C n.m.r. spectrum in that five signals were clearly discernible, arising from C-1,-2,-3,-9 and -13. This agreed entirely with our expectations since C-3,-9 and -13 arise from C-2 of mevalonate units in an eudesmane precurser while C-1 and -2 arise from an intact acetate fragment,  $^{2}$ C- $^{2}$ C, of a meva-

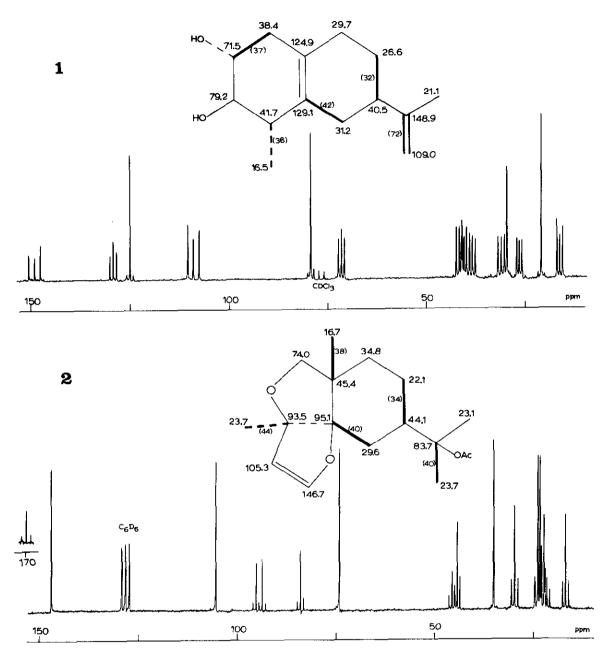


Fig. 1.  $^{13}\text{C}$  n.m.r. spectrum of  $^{3}\text{-}^{13}\text{Cx}$  isolated from the potato -  $\underline{\text{M}}$ . fructicola feeding experiment. The individual carbon shieldings are given in ppm from TMS and the  $J_{\text{C}}$  values in Hz (in parentheses) are shown for the pairs of carbons incorporated as intact acetate units.

Fig. 2.  $^{13}\text{C}$  n.m.r. spectrum of  $4^{-13}\text{C}_{X}$  isolated from the potato -  $\underline{6}$ .  $\underline{\text{cingulata}}$  experiment. The carbon shieldings are given in ppm from TMS and the J<sub>C-C</sub> values (in parentheses) are those measured for the intact acetate units, indicated by heavy bonds.

lonate unit which subsequently suffers scission to generate the phytuberin skeleton,  $^{4,6}$  as illustrated in Scheme 1.

## SCHEME 1

A more informative spectrum of  $\frac{4}{4}$  was nevertheless desired and for this purpose a similar feeding experiment was performed with  $\frac{610merella}{610merella}$  cingulata as the challenging fungus since this tends to give high yields of phytuberin. In the event, this led to generally lower incorporation levels but the potatoes (64 lb) afforded phytuberin (39 mg) which gave the  $^{13}\text{C}$  spectrum shown as Fig. 2. This shows the five intense signals lacking  $^{13}\text{C}$  satellites while each of the remaining absorption patterns consists of a central signal flanked by  $^{13}\text{C}$  satellites indicating that these arise from acetate units incorporated intact into the skeleton via mevalonate. The separation of each pair of satellite signals is the  $^{13}\text{C}-^{13}\text{C}$  coupling. With this information one can match the individual signals in pairs and readily complete the signal assignments for each carbon.

The potato - <u>G. cingulata</u> interaction also furnished, in addition to <u>I</u>, <u>2</u> and <u>3</u>, solavetivone  $^8$  (5) (17 mg) and dihydrolubimin  $^{9,10}$  (6) (<u>ca.</u> 120 mg), each of which exhibited the  $^{13}$ C

labelling patterns previously found for  $\underline{1}$  and  $\underline{2}$ , in agreement with expectation. The dihydro derivative  $\underline{6}$  probably arose at least in part through the metabolism of  $\underline{1}$  by  $\underline{6}$ . cingulata since this process is known to occur in pure culture.

Both feeding experiments afforded additional metabolites on which we shall report in the near future.

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